

Package: rACMEMEEV (via r-universe)

May 12, 2026

Title Multi-Variate Measurement Error Adjustment

Version 1.0.1

Description A methodology to perform multivariate measurement error adjustment using external validation data. Allows users to remove the attenuating effect of measurement error by incorporating a distribution of external validation data, and allows for plotting of all resultant adjustments. Sensitivity analyses can also be run through this package to test how different ranges of validity coefficients can impact the effect of the measurement error adjustment. The methods implemented in this package are based on the work by Muoka, A., Agogo, G., Ngesa, O., Mwambi, H. (2020):
<[doi:10.12688/f1000research.27892.1](https://doi.org/10.12688/f1000research.27892.1)>.

License MIT + file LICENSE

Encoding UTF-8

Roxygen list(markdown = TRUE)

RoxygenNote 7.3.2

Suggests knitr, rmarkdown, testthat (>= 3.0.0), BH, glmnet

Config/testthat/edition 3

Imports rjags, MCMCpack, ggplot2, reshape2, rstan, stats, utils, ggpubr, patchwork, bayesplot, doSNOW, foreach, snow, parallelly, dplyr

VignetteBuilder knitr

URL <https://github.com/westford14/rACME-MEEV>

BugReports <https://github.com/westford14/rACME-MEEV/issues>

Config/pak/sysreqs cmake make jags libicu-dev

Repository <https://westford14.r-universe.dev>

Date/Publication 2026-01-12 10:34:17 UTC

RemoteUrl <https://github.com/westford14/racme-meev>

RemoteRef HEAD

RemoteSha e3b1dfeb5893fe8e1a89a4de9fa33a1b4994b46c

Contents

acf_plots	2
acme_model	3
attenuation_matrix	4
create_model_string	5
create_modelling_data	6
create_stan_model_string	6
fisher_z_transform	7
generate_coefficient	8
multivariate_model	8
pipeline	10
plot_a_list	11
plot_covariates	12
sensitivity_analysis	13
standardize_with_return	14
traceplots	15
Index	17

acf_plots	<i>Create Auto Correlation Plots for Models</i>
-----------	---

Description

Create auto correlation plots for the models to test if there is any high autocorrelation in the sampling space, which would imply that sampler should be adjusted or the data needs to be adjusted.

Usage

```
acf_plots(input, columns, pre_model = FALSE, stan = FALSE)
```

Arguments

input	list List of the model output
columns	vector The target columns to assess
pre_model	bool If you are passing the pre-model
stan	bool If you are using the stan model

Value

plotted objects

Examples

```

columns <- c("fruit", "veg", "tobacco")
fruit_v_coef <- generate_coefficient(100, 0.3, 0.8, 0.95)
veg_v_coef <- generate_coefficient(100, 0.25, 0.75, 0.95)
tob_v_coef <- generate_coefficient(100, 0.4, 0.7, 0.95)
validity_coefficients <- c(fruit_v_coef, veg_v_coef, tob_v_coef)
data <- data.frame(
  list(
    "BMI" = rnorm(100, mean = 0, sd = 1),
    "fruit" = rnorm(100, mean = 0, sd = 1),
    "veg" = rnorm(100, mean = 0, sd = 1),
    "tobacco" = rnorm(100, mean = 0, sd = 1)
  )
)
output <- acme_model(data, columns)
lambda <- attenuation_matrix(
  output,
  columns,
  validity_coefficients,
)
model_output <- multivariate_model(
  "BMI ~ fruit + veg + tobacco",
  data = data,
  columns = columns,
  a_c_matrix = lambda$matrix,
  sds = lambda$sds,
  variances = lambda$variances,
  univariate = TRUE
)
acf_plots(model_output$naive, columns)

```

acme_model

Model the Data

Description

Using the methodology generated by Muoko et. al (see README for full citation), run the modelling with the JAGS sampler. This function accepts a number of different arguments, but defaulted arguments are assumed to be best for *most* systems. Obviously, if there is prior knowledge, this can be adjusted at the user's own discretion.

Usage

```

acme_model(
  data,
  columns,
  n_chains = 1,
  n_adapt_steps = 500,
  n_burn = 1000,

```

```

n_thin = 1,
n_steps = 10000,
seed = 42,
stan = FALSE
)

```

Arguments

data	data.frame	The data to format
columns	vector	The columns to target
n_chains	numeric	Number of chains to run, default = 1
n_adapt_steps	numeric	Number of adapt steps to run, default = 500
n_burn	numeric	Number of draws to burn at the start, default = 1000
n_thin	numeric	Thinning factor for the draws, default = 1
n_steps	numeric	The total number of draws to run, default = 10,000
seed	numeric	The random seed to set
stan	boolean	If you would like to use the experimental Stan backend

Value

List with all the appropriate things needed for modelling with JAGS

Examples

```

data <- data.frame(list("fruit" = c(1, 2), "veg" = c(3, 4)))
acme_model(data, names(data))

```

attenuation_matrix *Create Attenuation Contamination Matrix*

Description

From the output JAGS model, create the needed attenuation contamination matrix for further analysis. Please be aware that this is multivariate so a minimum of 3 columns is needed.

Usage

```
attenuation_matrix(model_output, columns, validity_coefficients, stan = FALSE)
```

Arguments

model_output	list	List of the mcmc summary and covariance matrix
columns	vector	Vector of the column names that are being assessed
validity_coefficients	vector	Vector of the validity coefficients
stan	boolean	If you are passing in a Stan backend pre-model

Value

List with the attenuation-contamination matrix and the standard deviations

Examples

```
columns <- c("fruit", "veg", "tobacco")
fruit_v_coef <- generate_coefficient(100, 0.3, 0.8, 0.95)
veg_v_coef <- generate_coefficient(100, 0.25, 0.75, 0.95)
tob_v_coef <- generate_coefficient(100, 0.4, 0.7, 0.95)
validity_coefficients <- c(fruit_v_coef, veg_v_coef, tob_v_coef)
data <- data.frame(
  list(
    "BMI" = rnorm(100, mean = 0, sd = 1),
    "fruit" = rnorm(100, mean = 0, sd = 1),
    "veg" = rnorm(100, mean = 0, sd = 1),
    "tobacco" = rnorm(100, mean = 0, sd = 1)
  )
)
output <- acme_model(data, columns)
attenuation_matrix(
  output,
  columns,
  validity_coefficients,
)
```

create_model_string *Define a Model that is JAGS Usable*

Description

Create the model string, save it to a temporary folder, and return back the location of the temporary model file for usage by JAGS later.

Usage

```
create_model_string()
```

Value

Full path to the model specification

Examples

```
create_model_string()
```

create_modelling_data *Data Preparation and Formatting for Modelling*

Description

Create the needed list that gets based to the JAGS backend for modelling. This should be a standard data.frame object that can then be standardized and reshaped into a format appropriate for modeling. Errors will be thrown if the data is not a class, or inherit from, a data.frame or if the specified columns do not exist in the data.frame.

Usage

```
create_modelling_data(data, columns)
```

Arguments

data	data.frame	The data to format
columns	vector	The columns to target

Value

List with all the appropriate things needed for modelling with JAGS

Examples

```
data <- data.frame(  
  list(  
    "BMI" = rnorm(100, mean = 0, sd = 1),  
    "fruit" = rnorm(100, mean = 0, sd = 1),  
    "veg" = rnorm(100, mean = 0, sd = 1),  
    "tobacco" = rnorm(100, mean = 0, sd = 1)  
  )  
)  
create_modelling_data(data, c("BMI", "fruit", "veg", "tobacco"))
```

create_stan_model_string

Define the Pre-Model Using Stan

Description

Create the model string, save it to a temporary folder, and return back the location of the temporary model file for usage by Stan later. This is experimental and has been validated, but caution should be used when utilizing the Stan backend as the pre-model.

Usage

```
create_stan_model_string()
```

Value

Full path to the model specification

Examples

```
create_stan_model_string()
```

fisher_z_transform *Perform the Fisher Z Transformation*

Description

Take a validity coefficient and perform the Fisher Z Transformation to approximate a normal distribution

Usage

```
fisher_z_transform(coefficient)
```

Arguments

coefficient numeric The validity coefficient to transform.

Value

Transformed validity coefficient

Examples

```
coef <- 0.3  
fisher_z_transform(coef)
```

generate_coefficient *Generate updated validity coefficient using Fisher Z Transformation*

Description

Using the Fisher Z Transformation, generate new validity coefficients based on proposed upper and lower boundaries. This new validity coefficient is based on transformed upper and lower boundaries as well as a fitted normal distribution to the se new proposed upper and lower boundaries.

Usage

```
generate_coefficient(n, lower_bound, upper_bound, interval = 0.95, seed = 42)
```

Arguments

n	the samples for the normal distribution
lower_bound	numeric the lower boundary of the validity coefficient
upper_bound	numeric the upper boudnary of the validity coefficient
interval	numeric the confidence interval to use (default 0.95)
seed	numeric the random seed to use

Value

Validity coefficient

Examples

```
n <- 1000
coef_upper <- 0.9
coef_lower <- 0.3
ci <- 0.95
generate_coefficient(n, coef_lower, coef_upper, ci)
```

multivariate_model *Model the Data with Multivariate Adjustment*

Description

Using the methodology generated by Muoko et. al (see README for full citation), run the modelling with the JAGS sampler. This model uses the calculated attenuation-contamination matrix to adjust the various covariates needed in the model. The function accepts the computed pre-model, the attenuation-contamination coefficient,, a model string, and standard deviations. There is an optional argument for also fitting the univariate model to run further comparisons between the naive, univariate, and multivariate models in later steps.

Usage

```

multivariate_model(
  formula,
  data,
  columns,
  a_c_matrix,
  n_burn = 1000,
  n_thin = 1,
  n_steps = 10000,
  seed = 42,
  b0 = 0,
  capital_b_0 = 1e-06,
  sampler = "Metropolis",
  c0 = 0.001,
  d0 = 0.001,
  univariate = FALSE,
  sds = NULL,
  variances = NULL
)

```

Arguments

formula	character	The model formula
data	data.frame	The data to model
columns	vector	The columns that are relevant
a_c_matrix	matrix	The attenuation-contamination matrix
n_burn	numeric	Number of draws to burn at the start, default = 1000
n_thin	numeric	Thinning factor for the draws, default = 1
n_steps	numeric	The total number of draws to run, default = 10,000
seed	numeric	The random seed to set
b0	numeric	The prior mean of the beta
capital_b_0	numeric	The precision of the beta
sampler	character	The sampler to use for the model
c0	numeric	Shape parameter for gamma
d0	numeric	Scale parameter for gamm
univariate	bool	Whether or not to run the univariate model, default = TRUE
sds	list	If you are running the univariate model, the listing of standard deviations needs to be applied
variances	list	If you are running the univariate model, the listing of variances need to be applied

Value

List with fitted models ready for further analysis

Examples

```

columns <- c("fruit", "veg", "tobacco")
fruit_v_coef <- generate_coefficient(100, 0.3, 0.8, 0.95)
veg_v_coef <- generate_coefficient(100, 0.25, 0.75, 0.95)
tob_v_coef <- generate_coefficient(100, 0.4, 0.7, 0.95)
validity_coefficients <- c(fruit_v_coef, veg_v_coef, tob_v_coef)
data <- data.frame(
  list(
    "BMI" = rnorm(100, mean = 0, sd = 1),
    "fruit" = rnorm(100, mean = 0, sd = 1),
    "veg" = rnorm(100, mean = 0, sd = 1),
    "tobacco" = rnorm(100, mean = 0, sd = 1)
  )
)
output <- acme_model(data, columns)
lambda <- attenuation_matrix(
  output,
  columns,
  validity_coefficients,
)
model_output <- multivariate_model(
  "BMI ~ fruit + veg + tobacco",
  data = data,
  columns = columns,
  a_c_matrix = lambda$matrix,
  sds = lambda$sds,
  variances = lambda$variances,
  univariate = TRUE
)

```

pipeline

Pipeline used for running a model start to finish.

Description

This is an unexported function that can run the entire pre-model, attenuation-contamination matrix, and model fitting from start to finish.

Usage

```
pipeline(data, formula, parameters, columns, stan = FALSE, seed = 42)
```

Arguments

data	data.frame	The input data
formula	character	The formula for the multivariate model
parameters	vector	The validity coefficients
columns	vector	The columns of the covariates

stan bool If you would like to run with the Stan backend
 seed numeric The random seed to use

Value

list The output parameters

Examples

```
data <- data.frame(
  list(
    "BMI" = rnorm(100, mean = 0, sd = 1),
    "fruit" = rnorm(100, mean = 0, sd = 1),
    "veg" = rnorm(100, mean = 0, sd = 1),
    "tobacco" = rnorm(100, mean = 0, sd = 1)
  )
)
parameters <- list(
  fruit = c(0.3, 0.55, 0.8),
  veg = c(0.25, 0.5, 0.75),
  tobacco = c(0.4, 0.55, 0.7)
)
grid <- expand.grid(parameters)
param_grid <- list()
for (i in seq_len(nrow(grid))) {
  name <- paste0("iteration_", i)
  param_grid[[name]] <- list(
    parameters = grid[i, ]
  )
}
output <- pipeline(
  data,
  "BMI ~ fruit + veg + tobacco",
  as.numeric(param_grid[[i]][["parameters"]]),
  c("fruit", "veg", "tobacco")
)
```

 plot_a_list

Custom Function to Use Patchwork

Description

Uses patchwork to make gridding easier for plotting the traceplots and acf plots for the Stan pre-model.

Usage

```
plot_a_list(plot_list, rows, columns)
```

Arguments

plot_list	list	List of plots
rows	numeric	Number of rows
columns	numeric	Number of columns

plot_covariates *Diagnostics for Models*

Description

Using the previously fit models, add the diagnostics for assessing the adjusted covariates based on the naive, univariate, and multivariate models.

Usage

```
plot_covariates(model_output, columns)
```

Arguments

model_output	list	List of the model output
columns	vector	The target columns to assess

Value

plotted objects

Examples

```
columns <- c("fruit", "veg", "tobacco")
fruit_v_coef <- generate_coefficient(100, 0.3, 0.8, 0.95)
veg_v_coef <- generate_coefficient(100, 0.25, 0.75, 0.95)
tob_v_coef <- generate_coefficient(100, 0.4, 0.7, 0.95)
validity_coefficients <- c(fruit_v_coef, veg_v_coef, tob_v_coef)
data <- data.frame(
  list(
    "BMI" = rnorm(100, mean = 0, sd = 1),
    "fruit" = rnorm(100, mean = 0, sd = 1),
    "veg" = rnorm(100, mean = 0, sd = 1),
    "tobacco" = rnorm(100, mean = 0, sd = 1)
  )
)
output <- acme_model(data, columns)
lambda <- attenuation_matrix(
  output,
  columns,
  validity_coefficients,
)
model_output <- multivariate_model(
```

```

    "BMI ~ fruit + veg + tobacco",
    data = data,
    columns = columns,
    a_c_matrix = lambda$matrix,
    sds = lambda$sds,
    variances = lambda$variances,
    univariate = TRUE
  )
  plot_covariates(model_output, columns)

```

sensitivity_analysis *Run a sensitivity analysis on the error adjustment*

Description

Create a sensitivity analysis based on a grid of validity coefficient parameters. The input of the parameter space should be a list with each key being the name of one of the covariates to vary and a vector of the parameter space to test the sensitivity. Be very careful because all combinations of the parameters will be tested so you can very easily run this for too long. Also please note that the parameters should be bound between 0 and 1, the theoretical limits of the validity coefficients. An example of this parameter grid is:

Usage

```

sensitivity_analysis(
  parameters,
  data,
  formula,
  columns,
  stan = FALSE,
  seed = 42
)

```

Arguments

parameters	list As described above
data	data.frame The data to use for the sensitivity analysis
formula	character The formula for the model
columns	vector The columns within the data for the covariates
stan	boolean Whether or not to run with the Stan backend
seed	numeric The random seed to set

Details

```
params <- list( fruit = c(0.1, 0.2, 0.3), veg = c(0.1, 0.2, 0.3), tobacco = c(0.1, 0.2, 0.3) )
```

But, again please note that this will then fit $3 * 3 * 3$ different models so the run time here can explode. Also parallel computation will be utilized here, but it is much more difficult to debug should errors arise. All this to say is: buyer beware.

Value

list with the means and the SDs of the parameters

Examples

```
columns <- c("fruit", "veg", "tobacco")
data <- data.frame(
  list(
    "BMI" = rnorm(5, mean = 0, sd = 1),
    "fruit" = rnorm(5, mean = 0, sd = 1),
    "veg" = rnorm(5, mean = 0, sd = 1),
    "tobacco" = rnorm(5, mean = 0, sd = 1)
  )
)
parameters <- list(
  fruit = c(0.3),
  veg = c(0.25),
  tobacco = c(0.4)
)
output_jags <- sensitivity_analysis(
  parameters,
  data,
  "BMI ~ fruit + veg + tobacco",
  columns
)
```

standardize_with_return

Standardize the data of 1-D vector

Description

Perform standardization of data on a 2-D dataframe type object. Standardization in this case refers to $(x - \text{mean}(x)) / \text{sd}(x)$ where X is a 1-dimensional vector.

Usage

```
standardize_with_return(data)
```

Arguments

data vector The vector to standardize

Value

List with the original standard deviation, mean, and the standardized data

Examples

```
data <- rnorm(100, mean = 0, sd = 1)
standardize_with_return(data)
```

 traceplots

Create Traceplots for the Parameters

Description

Create sampler traceplots to run diagnostics on whether or not the sampler converged when creating the posterior. Columns should always be in the order that they appear in the model.

Usage

```
traceplots(input, columns, pre_model = FALSE, stan = FALSE)
```

Arguments

input	list	List of the model output
columns	vector	The target columns to assess
pre_model	bool	If you are passing the pre-model
stan	bool	If the model is stan based

Value

plotted objects

Examples

```
columns <- c("fruit", "veg", "tobacco")
fruit_v_coef <- generate_coefficient(100, 0.3, 0.8, 0.95)
veg_v_coef <- generate_coefficient(100, 0.25, 0.75, 0.95)
tob_v_coef <- generate_coefficient(100, 0.4, 0.7, 0.95)
validity_coefficients <- c(fruit_v_coef, veg_v_coef, tob_v_coef)
data <- data.frame(
  list(
    "BMI" = rnorm(100, mean = 0, sd = 1),
    "fruit" = rnorm(100, mean = 0, sd = 1),
    "veg" = rnorm(100, mean = 0, sd = 1),
    "tobacco" = rnorm(100, mean = 0, sd = 1)
  )
)
output <- acme_model(data, columns)
lambda <- attenuation_matrix(
  output,
  columns,
  validity_coefficients,
)
```

```
model_output <- multivariate_model(  
  "BMI ~ fruit + veg + tobacco",  
  data = data,  
  columns = columns,  
  a_c_matrix = lambda$matrix,  
  sds = lambda$sds,  
  variances = lambda$variances,  
  univariate = TRUE  
)  
traceplots(model_output$naive, columns)
```

Index

acf_plots, 2
acme_model, 3
attenuation_matrix, 4

create_model_string, 5
create_modelling_data, 6
create_stan_model_string, 6

fisher_z_transform, 7

generate_coefficient, 8

multivariate_model, 8

pipeline, 10
plot_a_list, 11
plot_covariates, 12

sensitivity_analysis, 13
standardize_with_return, 14

traceplots, 15